

Syntheses of 22,23-Dihydro-1 α ,25-dihydroxyvitamin D₂ and Its 24*R*-Epimer, New Vitamin D₂ Derivatives

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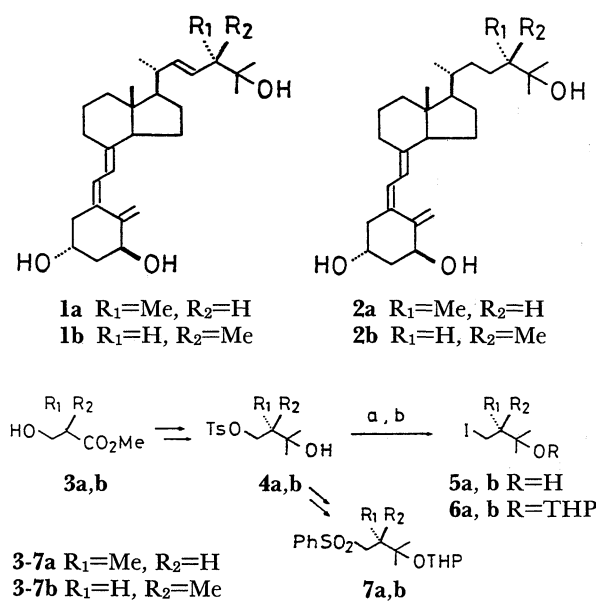
New 1 α ,25-dihydroxyvitamin D₂ derivatives (22,23-dihydro-1 α ,25-dihydroxyvitamin D₂ (**2a**) and its 24*R*-epimer (**2b**)), were synthesized by two procedures. 22-Oxo-5 α ,8 α -(4-phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-23,24-dinor-6-cholene-1 α ,3 β -diyl diacetate (**8**), obtainable readily from ergosterol, was converted to 22-phenylsulfonyl-5 α ,8 α -(4-phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-1 α ,3 β -bis(tetrahydropyranyloxy)-23,24-dinor-6-cholene (**14**) or 22-iodo-5 α ,8 α -(4-phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-1 α ,3 β -bis(tetrahydropyranyloxy)-23,24-dinor-6-cholene (**16**). Condensation of the C-22 sulfone (**14**) with (3*R*)-4-iodo-2,3-dimethyl-2-butanol THP ether, or the C-22 iodide (**16**) with (3*R*)-2,3-dimethyl-4-phenylsulfonyl-2-butanol THP ether followed by desulfonylation and successive deprotection gave 5,7-ergostadiene-1 α ,3 β ,25-triol (**21a**), which led to **2a** upon irradiation and subsequent thermal isomerization. Similarly, a 24*R*-epimer of **2a** was synthesized.

It is well established that vitamin D₃ and vitamin D₂ must be hydroxylated at the C-25 position in the liver, and subsequently at the C-1 α position in the kidney, before eliciting their physiological activity.¹⁾ Biological testing indicated that the activity of 1 α ,25-dihydroxyvitamin D₂ (**1a**) is similar to that of the corresponding vitamin D₃ derivative in mammals, though the former is 1/5—1/10 less active than the latter in birds.²⁾ Recently, DeLuca et al. reported that though 1 α -hydroxyvitamin D₂ is equally potent to 1 α -hydroxyvitamin D₃ regarding biological activity, the former is 5—10 times less toxic than the latter in rats.³⁾ These findings seem to be responsible for the difference of the side chain moiety between these "active" vitamin D₂ and vitamin D₃.

In order to study the functional importance of the 24-methyl group, the (24*R*)-1 α ,25-dihydroxyvitamin D₂^{4–6)} (**1b**) and (22*E*)- and (22*Z*)-dehydro-1 α -hydroxyvitamin D₃⁷⁾ were synthesized and their binding affinities investigated.⁸⁾ However, the influence of the C-22 double bond of these active vitamin D₂ derivatives on the physiological activity has not yet been clarified. These observations prompted us to study the effect of unsaturation at C-22,23 of 1 α ,25-dihydroxyvitamin D₂. In the present work, the syntheses of 22,23-dihydro-1 α ,25-dihydroxyvitamin D₂ (**2a**) and its 24*R*-epimer (**2b**), the new active vitamin D₂ derivatives, are described.

Our syntheses of **2a** and **2b** were achieved by two procedures. One was based on a condensation of the steroidal C-22 sulfone with optically active iodide derivatives to constitute the side chain part; the other was based on a coupling of the steroidal C-22 iodide with the corresponding side chain sulfone derivatives.

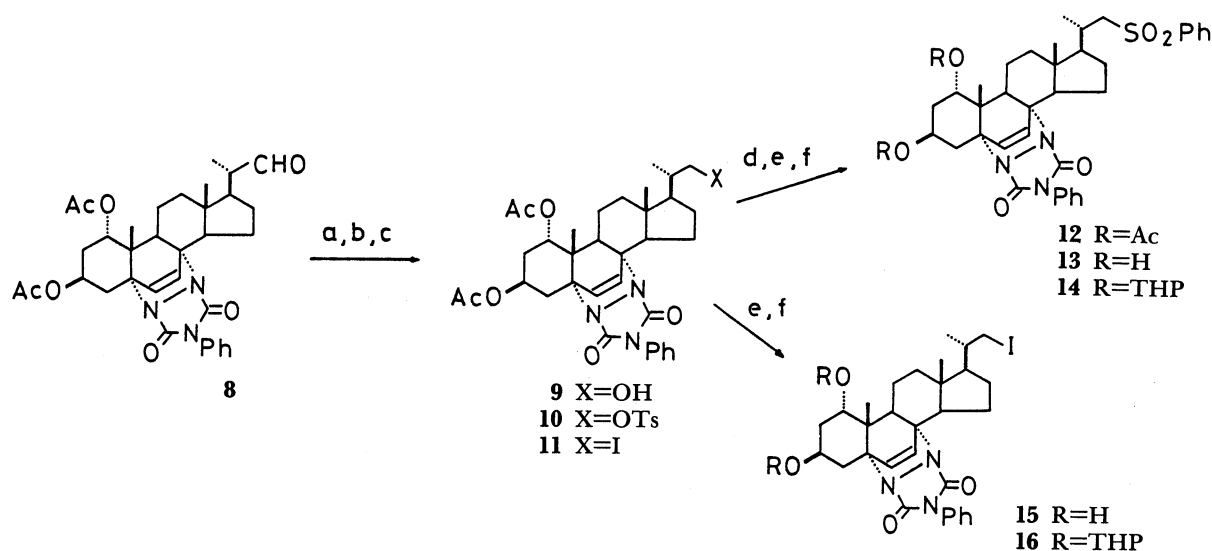
The side chain fragments (optically active iodides (**6a** and **6b**) and sulfones (**7a** and **7b**)) were synthesized, respectively, starting from methyl (*S*)- and (*R*)-3-hydroxy-2-methylpropionate (**3a** and **3b**) as follows (Scheme 1). The syntheses of **7a** and **7b** were reported previously.⁶⁾ The tosylate (**4a**), an intermediate of the



Scheme 1. a) NaI/acetone; b) DHP, PPTS.

sulfone (**7a**), was reacted with sodium iodide⁹⁾ in refluxing acetone to give the hydroxy iodide (**5a**) in 70% yield. The hydroxyl group of **5a** was protected as a tetrahydropyranyl (THP) ether in the usual manner to afford the iodide (**6a**) in 93% yield. Similarly, **6b** was prepared via the tosylate (**4b**).

The C-22 steroidal blocks, the C-22 sulfone (**14**) and the C-22 iodide (**16**) were synthesized as follows (Scheme 2). 22-Oxo-5 α ,8 α -(4-phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-23,24-dinor-6-cholene-1 α ,3 β -diyl diacetate (**8**), prepared by a method described in our previous paper,⁶⁾ seemed to be a good key intermediate for the preparation of **2a** and **2b** as the steroidal block. Thus, the aldehyde (**8**) was reduced by NaBH₄ in MeOH to afford the alcohol (**9**),¹⁰⁾ which was tosylated in the conventional manner to give the tosylate (**10**) in 78% yield from **8**. Treatment of **10** with sodium iodide in DMF was followed by a reaction with

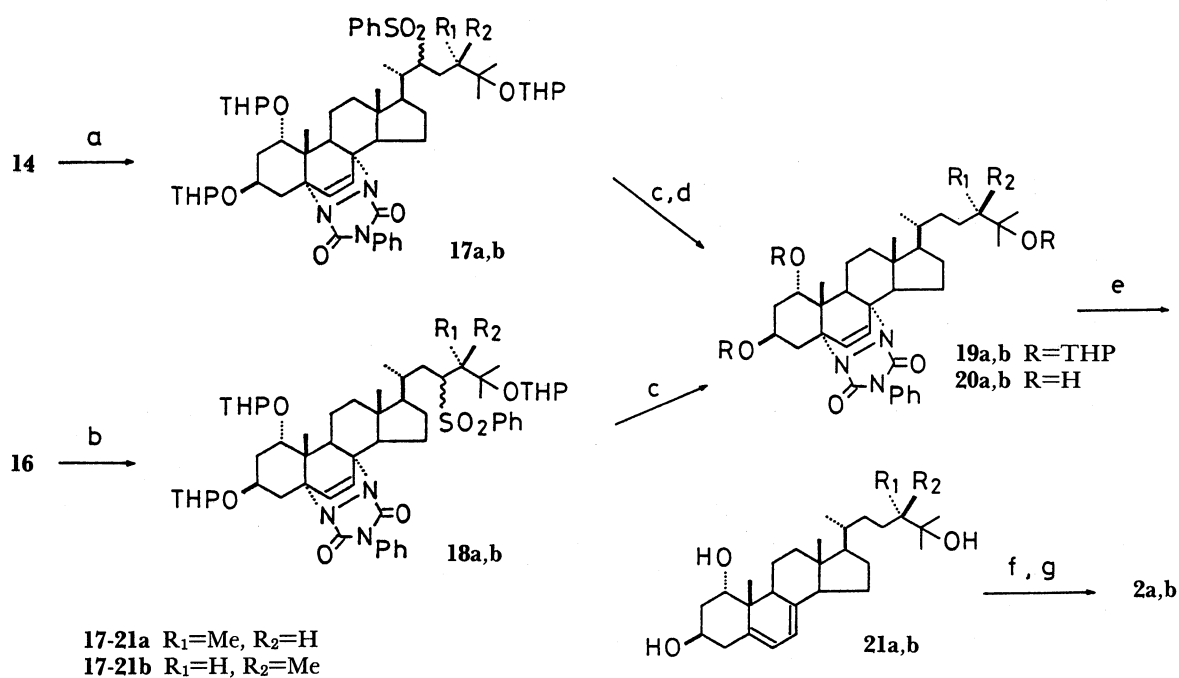


Scheme 2. a) NaBH₄/MeOH; b) *p*-TsCl, Py; c) NaI/DMF; d) PhSO₂Na/DMF; e) NaOH or KOH/MeOH; f) DHP, PPTS or *p*-TsOH.

sodium benzenesulfonate to yield the sulfone (**12**) in 82% yield.¹¹ The acetyl groups of **12** were converted to stable protective groups under the basic conditions employed in the following steps.¹⁰ Thus, the hydrolysis of **12** yielded the diol (**13**), which was protected as THP groups in the usual manner to give the steroidal C-22 sulfone (**14**) in 71% yield. Similarly, the conversion of the acetyl groups of **11** to the THP groups via the diol (**15**) gave the steroidal C-22 iodide (**16**) in 73% yield.

The next task was to introduce the side chain moiety (Scheme 3). Lithiation of the C-22 sulfone

(**14**) by butyllithium in the presence of hexamethylphosphoric triamide (HMPA) in THF at -20°C followed by the addition of the iodide (**6a**) gave the 22-phenylsulfonyl derivative (**17a**) in 44% yield (72% based on the consumed **14**) together with the recovered starting sulfone (**14**) in 38% yield. Reductive desulfonylation with sodium amalgam in buffered MeOH (Na₂HPO₄) and subsequent deprotection of the THP groups of the resulting **19a** provided the triol (**20a**) in 43% yield. The triol (**20a**) was treated with LiAlH₄ in refluxing THF to remove the triazoloedione protective group,¹² giving 5,7-ergostadiene-1 α ,3 β ,25-triol (**21a**)



Scheme 3. a) **6a** or **6b**, *n*-BuLi, HMPA/THF; b) **7a** or **7b**, *n*-BuLi, HMPA/THF; c) Na-Hg/MeOH (Na₂HPO₄); d) *p*-TsOH/EtOH; e) LiAlH₄/THF; f) *h\nu*; g) reflux/EtOH.

in 71% yield.

The introduction of a side chain was carried out by an alternative procedure. Condensation of the C-22 iodide (**16**) with the sulfone (**7a**) was achieved under the same conditions described above to yield the 23-phenylsulfonyl derivative (**18a**) in 60% yield. Desulfonylation of **18a** gave **19a** in 35% yield.

Irradiation of the 5,7-diene-1 α ,3 β ,25-triol (**21a**) with a high-pressure mercury lamp using an aq 1.5% KNO₃ solution as a filter followed by thermal isomerization of the resulting previtamin D in refluxing EtOH furnished crystalline 22,23-dihydro-1 α ,25-dihydroxyvitamin D₂ (**2a**) in 25% yield after purification by preparative HPLC.

Similarly, (24R)-5,7-ergostadiene-1 α ,3 β ,25-triol (**21b**) was prepared from C-22 sulfone (**14**) and iodide (**6b**) in place of **6a** in four steps with a 32% yield from **14**. Irradiation of **21b** and subsequent thermal isomerization of the resulting previtamin D gave crystalline (24R)-22,23-dihydro-1 α ,25-dihydroxyvitamin D₂ (**2b**) in 17% yield.

Since 22,23-dihydro-1 α ,25-dihydroxyvitamin D₂ (**2a**) and its 24R-epimer (**2b**) were prepared by two synthetic methods, the present study provides efficient routes to 1 α ,25-dihydroxyvitamin D derivatives having the C-22,23 single bond.

The biological activities of **2a** and **2b** will be reported elsewhere.

Experimental

All melting and boiling points are uncorrected. IR spectra were measured on a Jasco IR-810 spectrometer. ¹H NMR spectra were recorded with TMS as an internal standard and CDCl₃ as a solvent at 200 MHz on a JEOL JNM-FX 200 spectrometer, unless otherwise stated. Optical rotations were measured with CHCl₃ as a solvent on a Jasco DIP-370 polarimeter, unless otherwise stated. Mass spectra were recorded on a Hitachi M-80 spectrometer at 70 eV. Merck Kieselgel 60 (Art 7734, 70–230 mesh) or Merck Kieselgel 60 (Art 9385, 230–400 mesh) were used for SiO₂ column chromatography.

(3R)-4-Iodo-2,3-dimethyl-2-butanol THP Ether (6a). The tosylate (**4a**) was prepared from (S)-2,3-dimethyl-1,3-butanediol⁶ (6.40 g, 54.2 mmol) in the same manner as described previously.⁶ A solution of **4a** and sodium iodide (24.4 g, 0.16 mol) in acetone (180 ml) was stirred for 5 h at reflux temperature. After removing the acetone in vacuo, water was added to the residue and the mixture was extracted with ether. The ether solution was washed with a 10% Na₂S₂O₃ solution and brine, dried (MgSO₄), and concentrated in vacuo. The residue was distilled to give 8.26 g (70%) of **5a**: bp 69–71 °C/4 mmHg (1 mmHg \approx 133.322 Pa); n_D^{25} 1.5192; $[\alpha]_D^{25}$ –37.7° (*c* 1.98); IR (film) 3400, 1470, 1380, 1190, 1135, 1110, 950 cm⁻¹; ¹H NMR δ =1.11 (3H, d, *J*=6.8 Hz), 1.16 (3H, s), 1.26 (3H, s), 1.69 (1H, s), 1.87 (1H, m), 2.91 (1H, dd, *J*=9.5 and 10.5 Hz), 3.67 (1H, dd, *J*=9.5 and 7.2 Hz).

A solution of **5a** (7.73 g, 33.9 mmol), dihydropyran (5.70 g, 67.8 mmol) and pyridinium *p*-toluenesulfonate (PPTS) (0.85 g, 3.4 mmol) in dry CH₂Cl₂ (70 ml) was stirred for 3 h at

room temperature. The mixture was washed with a sat. NaHCO₃ solution and brine, dried (MgSO₄), and concentrated in vacuo. The residue was chromatographed over SiO₂ (130 g) eluting with hexane–ether (19:1) to give 9.88 g (93%) of **6a**: IR (film) 1470, 1390, 1375, 1200, 1130, 1075, 1035, 1025, 985 cm⁻¹. This was employed in the next step without further purification.

(3S)-4-Iodo-2,3-dimethyl-2-butanol THP Ether (6b). In the same manner as described for **5a**, (R)-2,3-dimethyl-1,3-butanediol⁶ (6.85 g, 58.1 mmol) was converted to 10.20 g (77%) of **5b**: bp 69–71 °C/4 mmHg; n_D^{25} 1.5190; $[\alpha]_D^{25}$ +38.7° (*c* 1.93). Its IR and ¹H NMR spectra were identical with those of **5a**.

In the same manner as described for **6a**, **5b** (8.60 g, 37.7 mmol) was converted to 10.88 g (92%) of **6b**. Its IR spectrum was identical with that of **6a**.

5 α ,8 α -(4-Phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-23,24-dinor-6-cholene-1 α ,3 β ,22-triyl 1 α ,3 β -Diacetate 22-*p*-Toluenesulfonate (10**).** To a stirred solution of **8** (9.50 g, 15.8 mmol) in MeOH (100 ml) was added NaBH₄ (0.30 g, 7.9 mmol) portionwise over 10 min at room temperature; the mixture was stirred for an additional 10 min. To the mixture was added AcOH (0.3 ml); the mixture was stirred for 10 min. After removal of MeOH in vacuo, water was added to the residue and the mixture was extracted with CHCl₃. The CHCl₃ solution was washed with water and brine, dried (MgSO₄), and concentrated in vacuo to give 9.50 g of crude alcohol (**9**).¹⁰ This was employed in the next step without further purification.

To a stirred, ice-cooled solution of **9** (9.50 g) in dry pyridine (45 ml) was added *p*-toluenesulfonyl chloride (4.50 g, 23.6 mmol); the mixture was stirred for 4 h at room temperature. To the reaction mixture was added a small amount of water and the mixture was stirred for 1 h at that temperature. The mixture was next poured into ice water and extracted with CHCl₃. The CHCl₃ solution was washed with water, 5% HCl, sat. NaHCO₃ solution and brine, dried (MgSO₄), and concentrated in vacuo. The residue was chromatographed over SiO₂ (250 g) eluting with hexane–EtOAc (1:1–1:2) to give 9.33 g (78% from **8**) of **10**: IR (KBr) 1750, 1700, 1600, 1505, 1400, 1245, 1180, 1030 cm⁻¹; ¹H NMR δ =0.80 (3H, s, 18-H₃), 1.01 (3H, d, *J*=6.6 Hz, 21-H₃), 1.05 (3H, s, 19-H₃), 2.01 (3H, s, Ac), 2.02 (3H, s, Ac), 2.44 (3H, s, –CH₃(tosyl)), 3.25 (1H, dd, *J*=13.4 and 5.6 Hz, 9-H), 3.73 (1H, dd, *J*=8.8 and 6.6 Hz, 22-H), 4.01 (1H, dd, *J*=8.8 and 2.4 Hz, 22-H), 5.09 (1H, m, 1-H), 5.87 (1H, m, 3-H), 6.33 and 6.41 (2H, ABq, *J*=8.3 Hz, 6-H and 7-H), 7.32–7.50 (8H, m, Ph), 7.77 (2H, d, *J*=8.1 Hz, Ph); MS *m/z* (rel intensity) 524 (M⁺–PTAD–AcOH; 3), 464 (58), 292 (43), 277 (16), 177 (62), 155 (100), 119 (78).

22-Iodo-5 α ,8 α -(4-phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-23,24-dinor-6-cholene-1 α ,3 β -diyl Diacetate (11**).** A solution of **10** (2.61 g, 3.44 mmol) and sodium iodide (2.57 g, 17.1 mmol) in dry DMF (20 ml) was stirred for 30 min at 80 °C. After cooling, the mixture was poured into water and extracted with CHCl₃. The CHCl₃ solution was washed with water, a 5% Na₂S₂O₃ solution and brine, dried (MgSO₄), and concentrated in vacuo. The residue was chromatographed over SiO₂ (50 g) eluting with hexane–EtOAc (3:2–1:1) to give 2.33 g (95%) of **11**: mp 173–174 °C (from hexane–EtOAc, rods); $[\alpha]_D^{25}$ –64.4° (*c* 1.12); IR (KBr) 1740, 1685, 1600, 1505, 1410, 1250, 1230, 1030 cm⁻¹; ¹H NMR δ =0.87 (3H, s, 18-H₃), 1.04 (3H, d, *J*=6.6 Hz, 21-H₃), 1.06

22-Phenylsulfonyl-5 α ,8 α -(4-phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-1 α ,3 β ,25-tris(tetrahydropyranyloxy)-6-ergostene (17a). To a stirred solution of **14** (3.50 g, 4.3 mmol) in dry THF (35 ml) was added successively butyllithium (1.5 mol dm⁻³ in hexane, 3.4 ml, 4.3 mmol) and dry HMPA (2.26 ml, 12.9 mmol) at -78°C under Ar, and the mixture was stirred for 20 min at -20°C . To the mixture was added a solution of **6a** (4.30 g, 12.9 mmol) in dry THF (12 ml); the mixture was stirred for 1.5 h at -20°C . The mixture was then poured into a sat. NH₄Cl solution and extracted with CHCl₃. The CHCl₃ solution was washed with brine, dried (MgSO₄), and concentrated in vacuo. The residue was chromatographed over SiO₂ (200 g) eluting with hexane-EtOAc (2:1–3:2–1:1). The earlier fraction gave 1.90 g (44%; 72% based on the consumed **14**) of **17a**, and the latter fraction gave 1.34 g (38%) of the recovered **14**: **17a**, IR (KBr) 1750, 1695, 1605, 1505, 1400, 1150, 1130, 1075, 1030, 985 cm⁻¹; ¹H NMR δ =3.05 (1H, m, 22-H), 3.22 (1H, m, 9-H), 3.48 (3H, m, CH₂(THP)), 3.69 (1H, m, 1-H), 3.93 (3H, m, CH₂(THP)), 4.78 (3H, m, CH(THP)), 4.93 (1H, m, 3-H), 6.33 (2H, m, 6-H and 7-H), 7.3–7.9 (10H, m, Ph); MS m/z (rel intensity) 552 (M⁺-PTAD-DHP \times 3-H₂O; 16), 534 (21), 177 (60), 119 (100).

(24R)-22-Phenylsulfonyl-5 α ,8 α -(4-phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-1 α ,3 β ,25-tris(tetrahydropyranyloxy)-6-ergostene (17b). In the same manner as described for **17a**, **14** (2.55 g, 3.1 mmol) and **6b** (2.93 g, 9.4 mmol) was converted to 1.43 g (46%; 84% based on the consumed **14**) of **17b** together with 1.17 g (46%) of the recovered **14**: **17b**, IR (KBr) 1750, 1695, 1605, 1505, 1400, 1150, 1130, 1080, 1030, 985 cm⁻¹; ¹H NMR δ =3.18 (2H, m, 9-H and 22-H), 3.48 (3H, m, CH₂(THP)), 3.70 (1H, m, 1-H), 3.93 (3H, m, CH₂(THP)), 4.80 (3H, m, CH(THP)), 4.92 (1H, m, 3-H), 6.32 (2H, m, 6-H and 7-H), 7.3—7.9 (10H, m, Ph).

23-Phenylsulfonyl-5 α ,8 α -(4-phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-1 α ,3 β ,25-tris(tetrahydropyranyloxy)-6-ergostene (18a). To a stirred solution of **7a** (326 mg, 1.0 mmol) in dry THF (3 ml) was added successively butyllithium (1.5 mol dm⁻³ in hexane, 0.67 ml, 1.0 mmol) and dry HMPA (0.17 ml, 1.0 mmol) at -78 °C under Ar; the mixture was stirred for 20 min at -20 °C. To the mixture was next added a solution of **16** (400 mg, 0.50 mmol) in dry THF (4 ml). After stirring for 2 h at -20 °C, the mixture was stirred for 2 h at room temperature, poured into a sat. NH₄Cl solution, and extracted with CHCl₃. The CHCl₃ solution was washed with brine, dried (MgSO₄), and concentrated in vacuo. The residue was chromatographed over SiO₂ (20 g) eluting with hexane-EtOAc (4:1) to give 317 mg (64%) of **18a**: IR (KBr) 1750, 1695, 1600, 1500, 1400, 1140, 1125, 1030 cm⁻¹; ¹H NMR δ =3.1—3.6 (5H, m, 9-H, 23-H and CH₂(THP)), 3.67 (1H, m, 1-H), 3.87 (3H, m, CH₂(THP)), 4.75 (3H, m, CH(THP)), 4.95 (1H, m, 3-H), 6.33 (2H, m, 6-H and 7-H), 7.3—7.9 (10H, m, Ph); MS *m/z* (rel intensity) 570 (M⁺-PTAD-DHP \times 3; 3), 552 (4), 534 (4), 177 (57), 119 (100).

5 α ,8 α -(4-Phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-1 α ,3 β ,25-tris(tetrahydropyranyloxy)-6-ergostene (19a). (a) (from **17a**) To a solution of **17a** (1.20 g, 1.2 mmol) in MeOH saturated with Na₂HPO₄ (120 ml) was added sodium amalgam (5%, 16.6 g, 36.0 mmol), and the mixture was stirred for 16 h at room temperature. After removal of MeOH in vacuo from the supernatant, water was added to the residue and the mixture extracted with CHCl₃. The CHCl₃ solution was washed with water and brine, dried (MgSO₄), and concentrated in vacuo. The residue was chromatographed over SiO₂ (20 g) eluting with hexane-EtOAc (3:1—2:1) to give 0.51 g (50%) of **19a**: IR (KBr) 1750, 1700, 1605, 1505, 1400, 1135, 1080, 1030, 985 cm⁻¹; ¹H NMR δ =3.22 (1H, m, 9-H), 3.47 (3H, m, CH₂(THP)), 3.70 (1H, m, 1-H), 3.92 (3H, m, CH₂(THP)), 4.78 (3H, m, CH(THP)), 4.93 (1H, m, 3-H), 6.37 (2H, m, 6-H and 7-H), 7.3—7.5 (5H, m, Ph); MS *m/z* (rel intensity) 598 (M⁺-PTAD-DHP; 4), 580 (1), 412 (80), 239 (18), 177 (85), 119 (100).

(b) (from **18a**) In the same manner as described above, **18a** (300 mg, 0.30 mmol) was converted to 90 mg (35%) of **19a**. Its IR and ¹H NMR spectra were identical with those described above.

(24R)-5 α ,8 α -(4-Phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-1 α ,3 β ,25-tris(tetrahydropyranyloxy)-6-ergostene (19b). In the same manner as described for **19a**, **17b** (1.17 g, 1.7 mmol) was converted to 0.78 g (53%) of **19b**: IR (KBr) 1750, 1695, 1600, 1505, 1395, 1130, 1075, 1025, 985 cm⁻¹; ¹H NMR δ =3.20 (1H, m, 9-H), 3.47 (3H, m, CH₂(THP)), 3.70 (1H, m, 1-H), 3.92 (3H, m, CH₂(THP)), 4.78 (3H, m, CH(THP)), 4.93 (1H, m, 3-H), 6.37 (2H, m, 6-H and 7-H), 7.3—7.5 (5H, m, Ph).

5 α ,8 α -(4-Phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-6-

ergostene-1 α ,3 β ,25-triol (20a). A solution of **19a** (0.51 g, 0.60 mmol) and *p*-TsOH·H₂O (23 mg, 0.12 mmol) in 95% EtOH (5 ml) was stirred for 4 h at 80 °C. After removal of EtOH in vacuo, brine was added to the residue and the mixture was extracted with CHCl₃. The CHCl₃ solution was dried (MgSO₄) and concentrated in vacuo. The residue was chromatographed over SiO₂ (25 g) eluting with CHCl₃-EtOAc (1:3) and then EtOAc to give 0.31 g (85%) of **20a**: mp 211—214 °C (from EtOAc, rods); [α]_D²⁵ -97.7° (c 0.31); IR (KBr) 3530, 3460, 1745, 1680, 1505, 1410, 1320, 1150, 1035 cm⁻¹; ¹H NMR δ =0.81 (3H, s, 18-H₃), 0.88 (3H, d, *J*=7.1 Hz, 28-H₃), 0.92 (3H, s, 19-H₃), 0.94 (3H, d, *J*=6.4 Hz, 21-H₃), 1.14 and 1.15 (6H, each s, 26-H₃ and 27-H₃), 3.12 (1H, dd, *J*=15.6 and 6.1 Hz, 9-H), 3.85 (1H, m, 1-H), 4.88 (1H, m, 3-H), 6.25 and 6.41 (2H, ABq, *J*=8.5 Hz, 6-H and 7-H), 7.3—7.4 (5H, m, Ph); MS *m/z* (rel intensity) 430 (M⁺-PTAD; 13), 412 (12), 394 (11), 251 (17), 199 (41), 119 (100). Found: *m/z* 430.3444. Calcd for C₂₈H₄₆O₃ (M⁺-PTAD): M, 430.3449.

(24R)-5 α ,8 α -(4-Phenyl-3,5-dioxo-1,2,4-triazolidine-1,2-diyl)-6-ergostene-1 α ,3 β ,25-triol (20b). In the same manner as described for **20a**, **19b** (0.77 g, 0.90 mmol) was converted to 0.45 g (83%) of **20b**: mp 216—218 °C (from EtOAc-MeOH, needles); [α]_D²⁵ -79.0° (c 0.36); IR (KBr) 3520, 1745, 1680, 1505, 1410, 1325, 1155, 1035 cm⁻¹; ¹H NMR δ =0.82 (3H, s, 18-H₃), 0.87 (3H, d, *J*=6.8 Hz, 28-H₃), 0.93 (3H, s, 19-H₃), 0.95 (3H, d, *J*=6.7 Hz, 21-H₃), 1.15 and 1.17 (6H, each s, 26-H₃ and 27-H₃), 3.15 (1H, dd, *J*=16.4 and 7.1 Hz, 9-H), 3.88 (1H, m, 1-H), 4.90 (1H, m, 3-H), 6.27 and 6.42 (2H, ABq, *J*=6.8 Hz, 6-H and 7-H), 7.30—7.42 (5H, m, Ph); MS *m/z* (rel intensity) 430 (M⁺-PTAD; 8), 251 (15), 177 (44), 119 (100). Found: C, 71.39; H, 8.52; N, 7.01%. Calcd for C₃₆H₅₁N₃O₅: C, 71.37; H, 8.49; N, 6.94%.

5,7-Ergostadiene-1 α ,3 β ,25-triol (21a). To a suspension of LiAlH₄ (0.40 g) in dry THF (30 ml) was added a solution of **20a** (0.44 g, 0.73 mmol) in dry THF (10 ml); the mixture was stirred for 1.5 h at reflux temperature. Then to the ice-cooled mixture was added successively water (0.4 ml), a 10% NaOH solution (0.4 ml) and water (1.2 ml); the mixture was stirred for 30 min at room temperature. After addition of MgSO₄, the mixture was stirred for an additional 30 min and filtered through Celite. The filtrate was concentrated in vacuo; thus, the residue was recrystallized from EtOH-THF to give 0.22 g (71%) of **21a** as needles: mp 228—231 °C; [α]_D²⁵ -89° (c 0.11, THF); IR (KBr) 3520, 3360, 1655, 1605, 1465, 1380, 1135, 1070, 1045 cm⁻¹; ¹H NMR (DMSO-*d*₆+CDCl₃) δ =0.60 (3H, s, 18-H₃), 0.84 (3H, d, *J*=6.6 Hz, 28-H₃), 0.85 (3H, s, 19-H₃), 0.95 (3H, d, *J*=6.1 Hz, 21-H₃), 1.07 and 1.08 (6H, each s, 26-H₃ and 27-H₃), 3.62 (1H, m, 1-H), 3.91 (1H, m, 3-H), 5.30 (1H, m, 7-H), 5.56 (1H, m, 6-H); MS *m/z* (rel intensity) 430 (M⁺; 55), 412 (85), 394 (31), 251 (40), 197 (64), 157 (100), 145 (68); UV (EtOH) 282 nm (ϵ 9200). Found: C, 77.59; H, 10.70%. Calcd for C₂₈H₄₆O₃: C, 78.09; H, 10.77%.

(24R)-5,7-Ergostadiene-1 α ,3 β ,25-triol (21b). In the same manner as described for **21a**, **20b** (0.45 g, 0.74 mmol) was converted to 0.28 g (86%) of **21b**: mp 154—157 °C (from EtOH, rods); [α]_D²⁵ -17° (c 0.12, MeOH); IR (KBr) 3400, 1655, 1605, 1465, 1385, 1155, 1055 cm⁻¹; ¹H NMR δ =0.63 (3H, s, 18-H₃), 0.88 (3H, d, *J*=6.6 Hz, 28-H₃), 0.95 (3H, d, *J*=6.1 Hz, 21-H₃), 0.95 (3H, s, 19-H₃), 1.16 and 1.17 (6H, each s, 26-H₃ and 27-H₃), 3.78 (1H, m, 1-H), 4.08 (1H, m, 3-H), 5.40 (1H, m, 7-H), 5.73 (1H, m, 6-H); MS *m/z* (rel intensity) 430 (M⁺; 32), 412 (20), 394 (18), 251 (35), 197 (64), 157 (100), 145 (65). Found: *m/z* 430.3443. Calcd for C₂₈H₄₆O₃: M, 430.3449.

22,23-Dihydro-1 α ,25-dihydroxyvitamin D₂ (2a). A solution of **21a** (100 mg, 0.23 mmol) in ether-THF (19:1, 1000 ml) was irradiated for 3 min under Ar at water-cooled temperature with a high-pressure mercury lamp (Ushio, UM-452) using 1.5% KNO₃ solution as a filter. The mixture was concentrated in vacuo. A solution of the residue containing the previtamin D in EtOH (30 ml) was stirred for 1 h at reflux temperature under Ar, and then concentrated in vacuo. The residue was chromatographed on HPLC (Merck, LiChrosorb® Si60 (7 μ m), 25×250 mm) eluting with 6% MeOH-CH₂Cl₂ (6.0 ml min⁻¹) to give 25 mg (25% from **21a**) of crystalline **2a**. This was recrystallized from hexane-CH₂Cl₂ to give **2a** as needles: mp 93–95 °C; [α]_D²⁵ +33° (c 0.15, EtOH); ¹H NMR δ =0.54 (3H, s, 18-H₃), 0.90 (3H, d, *J*=6.8 Hz, 28-H₃), 0.94 (3H, d, *J*=5.9 Hz, 21-H₃), 1.15 and 1.17 (6H, each s, 26-H₃ and 27-H₃), 4.23 (1H, m, 3-H), 4.43 (1H, m, 1-H), 5.00 (1H, narrow m, 19-H), 5.33 (1H, narrow m, 19-H), 6.02 (1H, d, *J*=11.2 Hz, 7-H), 6.38 (1H, d, *J*=11.2 Hz, 6-H); MS *m/z* (rel intensity) 430 (M⁺; 8), 412 (10), 394 (11), 285 (6), 251 (5), 134 (100), 105 (34); UV (EtOH) 265 nm (ϵ 16900).

(24R)-22,23-Dihydro-1 α ,25-dihydroxyvitamin D₂ (2b). A solution of **21b** (100 mg, 0.23 mmol) in ether (1000 ml) was irradiated for 3 min under the same conditions as described above, and then concentrated in vacuo. The residue was chromatographed on HPLC (Merck, LiChrosorb® Si60 (7 μ m), 25×250 mm) eluting with 6% MeOH-CH₂Cl₂ (6.0 ml min⁻¹) to give 25.0 mg (25%) of the previtamin D: ¹H NMR δ =0.70 (3H, s, 18-H₃), 0.88 (3H, d, *J*=6.8 Hz, 28-H₃), 0.95 (3H, d, *J*=6.1 Hz, 21-H₃), 1.17 (6H, br s, 26-H₃ and 27-H₃), 1.77 (3H, s, 19-H₃), 4.06 (1H, m, 3-H), 4.20 (1H, m, 1-H), 5.50 (1-H, m, 9-H), 5.78 and 5.92 (2H, ABq, *J*=12.2 Hz, 6-H and 7-H).

A solution of the previtamin D (25.0 mg) in EtOH (15 ml) was stirred for 1 h at reflux temperature, and concentrated in vacuo. The residue was chromatographed on HPLC under the same conditions as mentioned above to give 16.7 mg (67%; 17% from **21b**) of crystalline **2b**. This was recrystallized from hexane-CH₂Cl₂ to give **2b** as rods: mp 172–

174 °C; [α]_D²⁵ +63° (c 0.11, EtOH); ¹H NMR δ =0.54 (3H, s, 18-H₃), 0.88 (3H, d, *J*=6.8 Hz, 28-H₃), 0.93 (3H, d, *J*=6.1 Hz, 21-H₃), 1.16 and 1.17 (6H, each s, 26-H₃ and 27-H₃), 4.23 (1H, m, 3-H), 4.44 (1H, m, 1-H), 5.01 (1H, narrow m, 19-H), 5.33 (1H, narrow m, 19-H), 6.02 (1H, d, *J*=11.2 Hz, 7-H), 6.38 (1H, d, *J*=11.2 Hz, 6-H); MS *m/z* (rel intensity) 430 (M⁺; 5), 412 (11), 394 (18), 285 (5), 251 (5), 134 (100), 105 (32); UV (EtOH) 265 nm (ϵ 17600).

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